

Detection and Recovery of E. coli O157:H7 in Alfalfa sprouts by Real-time PCR Combined with Immunomagnetic Separation With and Without an Acid Treatment

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SUMMARY

E. coli O157:H7 has been implicated in foodborne disease outbreaks with alfalfa sprouts. However, detection in sprouts by standard cultural methods can be difficult due to the high backgro croflora. This study evaluated an improved procedure for rapid detection and isolation of E. coli O157:H7 from sprout samples.

Alfalfa sprouts were artificially contaminated with E. coli O157:H7 at low (~0.2 cfu/g) and high (~2 cfu/g) levels and enriched in modified buffered peptone water + pyruvate (mBPWp) with cefsulodin and vancomycin at 42°C with shaking. After 5h and 54h, the target organisms were concentrated by IMS using PATHATIRX^{TW} or Dynabeads⁴⁷ MAX *E*. *coli* (0157). MS beads were screened by real-time PCR for simultaneous detection of stx1, stx2 and uidA genes using the SmartCycler. Additionally, broth cultures and IMS beads were streaked onto selective agar plates (Rainbow[#] agar, R&F[#] E. coli O157 Chromogenic medium and TC-SMAC agar) for isolation of E. coli O157:H7. Both broth culture and IMS beads were also acid treaded to improve upon cultural recovery.

After 5 h enrichment and PATHATRIXTM IMS, E. coli O157:H7 was detected by real-time PCR in After 3 in entrement and PATIFATKA* 1005, E. COU (3)7, r11 wis detected by fear-line FCA. 32/5 samples incoulated at the low level, isolated from 725 incoulated samples on selective agars, but from 22/25 following acid treatment of the beads. *E. coli* (0157 was detected in 22/25 samples isolated from 02/25 following MS with the Dynabead* MAX *E. coli* (0157 system, but from 15/25 following acidification of the beads. After 24h enrichment, cultural recovery was improved for enrichment broths and both types of IMS beads following acid treatment of the samples. Acidification of E. coli O157 enrichment broths and IMS beads improved isolation by eliminating competing organisms that make isolation difficult.

INTRODUCTION

A procedure for the enrichment of Shiga taxin producing E *coli* in foods using selective enrichment of samples in modified Buffered Peptone Water (mBPWp) (Weagant & Bound, 2001) was combined with a rapid, real-ime PCR assay specific for srat, 1, arX, and the +93 *iidA* mutation to screen for the sence of E. coli O157:H7 in foods (Jinneman et al., 2003: Yoshitomi et al., 2003) and is canable of detecting < 1 CFU/g of STEC in foods.

Enrichment in mBPWp at 42°C under shaking with antibiotics (cefsulodin and vancomycin) followed Enrichment introl w far at 2 Curice standing with automote (existication and variation) politicate by IMS was shown to be an effective method to detect EHE in a fallane sportus (Wenddoon et al., 2008). However, alfalfa sprout enrichment broths present a challenge in isolating EHEC, even with the use of IMS, because competing microorganisms that grow in the enrichment, are co-extracted with *E*. coll 0157 by IMS and subsequently grow on the selective agar plates (Weagant et al. 2008). We evaluated the optimized enrichment with alfalfa sprouts artificially contaminated with five EHEC strains and continued to encounter difficulty in recovering the inoculated EHEC on the selective plates (Jinneman et al., 2009). An acid treatment of IMS beads and enrichment broth cultures was found to improve recovery of EHEC from artificially contaminated cilanto (Fedio, et al, 2009).

The Pathatrix™ (Matrix Microscience, CO, USA), and the Invitrogen Bead Retriever with Dynabeads The Fanal the "(matter informatics, O_1 , O_2), and the invitingent beam relative with phases MAX E, cold O157 were used for detection and isolation of E, cold 0157. E to HMS system uses magnetic beads coated with antibodies to capture the target bacteria. The beads with captured target organisms are ready for further use in PCR detection or cultural isolation methods. In addition, an acidification treatment was employed to improve cultural recovery on selective agars.

MATERIALS AND METHODS

Bacterial Strain

Five E. coli O157:H7 strains were used for inoculation of the alfalfa sprouts: E. coli O157:H7 ATCC 35150 (stx1, stx2), ATCC 43895 (stx1, stx2), ATCC 43894 (stx1, stx2), ATCC 43889 (stx2 only), and TTT (srk1 only). Stationary phase cultures were diluted in Butterfield's Phosphate Buffer and used to inoculate the alfalfa sprouts.

Preparation of Alfalfa sprouts samples

Treparation or neural process samples is a process of the set of

Enrichment procedury

Enrement procedure Modified Buffered Peptone Water with pyruate (mBPWp) was used to enrich *E. coli* O157:H7 in the sprout samples. For each 25 g sample, 225 ml of pre-warmed mBPWp was added. Samples were stomached for 2 min and antibiotics were added. A ntibiotics: (final concentration in mBPWp): Cefsulodin (10mg/l), Vancomycin (8mg/l). S amples were incubated at 42°C with shaking (100 rpm) for 5h or 24h

After 5 h enrichment, samples were immunoconcentrated by the Pathatrix and Bead Retriever system: After 3 in encimienti, suppose were immunoconcentrated by the randomized and been kettered systems. The HdS backs were plated onto selective media or acid treated prior to plaing as outlined in Figure 1. A portion of the beads were also used for real time PCR detection with the primers as probes listed in Table 1 on the Smart Cycler II as shown in Figure 2. After 24 he nerichment profit samples and IMS backs were used for PCR and both non-acid treated and acid treated samples were plated onto selective agars



Figure 1. Procedure for analyzing enrichment cultures by real time PCR, Pathatrix IMS and Invitrogen IMS for detection of E. coli O157:H7 and isolation on selective as



Bead Retriever System



Figure 2. Real-time PCR detection of E. coli O157:H7 stx1, stx2, and +93 uidA markers. Smart cycle II equipment (left) and graph of amplification (right).



Table 3. Detection of E. coli O157:H7 in alfalfa sprouts artificially contaminated at a high level (1-3 cfu/g), enriched in mBPWp with cefsulodin (10mg/L) and vancomycin (8mg/L) and incubated with shaking at 42°C for 5h. D etection by real time PCR, and cultural isolation with or without IMS and an acid treatment

		-		Pathatrix b	eads		Dynaheads@www.X.F.coli 0157						
E.coli 0157 (inoculum level)	Incubation Time	PCR	Cultural isolation	Rainbow	TC-SMAC	R & F	PCR	Cultural isolation	Rainbow	TC-SMAC	R&F		
ATCC 35150 (1.8 cfu/g)	5h	5/5	4/5	1/5	3/5	1/5	5/5	2/5	2/5	0/5	0/5		
ATCC 43894 (2.0 cfu/g)	5h	5/5	5/5	2/5	5/5	3/5	5/5	2/5	0/5	1/5	1/5		
ATCC 43895 (1.8 cfu/g)	5h	5/5	5/5	0/5	5/5	1/5	5/5	0/5	0/5	0/5	0/5		
ATCC 43889 (2.7cfu/g)	5h	4/5	2/5	0/5	2/5	0/5	5/5	0/5	0/5	0/5	0/5		
TT7 (1.8 cfu/g)	5h	5/5	4/5	4/5	3/5	0/5	5/5	2/5	2/5	1/5	0/5		
Total		24/25	20/25	7/25	18/25	5/25	25/25	6/25	4/25	2/25	1/25		
Acid Treated													
ATCC 35150 (1.8 cfu/g)	5h		5/5	5/5	3/5	4/5		5/5	5/5	4/5	0/5		
ATCC 43894 (2.0 cfu/g)	5h		5/5	5/5	4/5	4/5		5/5	4/5	5/5	5/5		
ATCC 43895 (1.8 cfu/g)	5h		5/5	4/5	5/5	5/5		4/5	1/5	3/5	2/5		
ATCC 43889 (2.7ctu/g)	5h		5/5	5/5	0/5	0/5		4/5	3/5	4/5	2/5		
TT7 (1.8 cfu/g)	Sh		5/5	5/5	4/5	45		5/5	5/5	4/5	4/5		
Total			25/25	24/25	16/25	17/25		23/25	18/25	20/25	13/25		



Figure 4. Isolation of E. coli O157 from alfalfa sprouts on Rainbow agar. Top row: 24h enrichment broth streak plate (10 µ1 of undiluted enrichment broth) and spread plate (50 ul of 10-3 and 10-5 dilution), p ottom row Spread plates of acid treated alfalfa sprout nrichment broth (50 µ1 of undiluted, 10⁻¹ and 10⁻² dilutions of acid treated enrichment

coli O157 (right).

Figure 5. Isolation of E. coli O157 from Figure 6. Isolation of E. coli O157 from alfalfa alfalfa sprouts on TC-SMAC agar. T op rov 24h enrichment broth streak plate (10 µ1 of undiluted enrichment broth) and spread plate (50 µl of 10⁻³ and 10⁻⁵ dilution). Bottom row Spread plates of acid treated alfalfa sprout enrichment broth (50 µ1 of undiluted, 10and 10-2 dilutions of acid treated enrichment



Figure 8. Isolation of E. coli O157 from alfalfa sprout Figure 7. Isolation of E coli O157 from alfalfa sprout Figure 7. Isolation of E. coli O157 from attain spro enrichment broths following Pathatrix IMS. Top row: Pathatrix IMS bead streak plates (15 μ 1 of IMS beads from 24 h enrichment broth) on Rainbow agar (left), TC SMAC (center) and R&F Chromogenia medium for E. coli O157 (right). Bottom row: Acid treated Pathatrix IMS beads on Rainbow agar (left), TC SMAC (center) and R&F Chromogenic medium for E. R&F agar(right). commendation by the U.S. Food and Drug Administration Reference to any commercial materials, equipment, or process does not in any way constitute approval, endorsement, or re-

Figure 8. Isolation of *E. coli* OTS/ from attalta sprou-enrichment broths following Invitrogen IMS. Top row: Dynabeads[#] MAX anti-*E. coli* OTS7 beads streak plates (15 µl of IMS beads from 24 h enrichment broth) on Rainbow agar (left), TC SMAC (center) and R&F Chromogenic medium for *E. coli* O157 (right). Bottom row: Acid treated Dynabeads[®] MAX anti-*E. coli* O157 beads on Rainbow agar (left), TC SMAC (center) and

sprouts on R & F Chromogenic medium for E.

coli O157. T op row: 24h enrichment broth streak plate (10 µl of undiluted enrichment broth) and spread plate (50 µl of 10-3 and 10-5 dilution).

Bottom row: Spread plates of acid treated alfalfa

sprout enrichment broth (50 µ1 of undiluted, 10-

and 10-2 dilutions of acid treated enrichment)

E coli orra	he autorite a		Broth				Pathatrix beads						Dynabeads® M AX E.coli 0157					
(inoculum level)	Time	PCR	Cultural isolation	Rainbow	TC-SMAC	R & F	PCR	Cultural isolation	Rainbow	TC-SMAC	R&F	PCR	Cultural isolation	Rainbow	TC-SMAC	R&F		
ATCC 35150 (1.8 ctu/g)	24h	2/5	5/5	5/5	5/5	3/5	4/5	5/5	5/5	5/5	4/5	4/5	5/5	5/5	5/5	5/5		
ATCC 43894 (2.0 ctu/g)	24h	5/5	3/5	1/5	1/5	1/5	4/5	4/5	3/5	4/5	4/5	5/5	2/5	2/5	0/5	1/5		
ATCC 43895 (1.8 ctu/g)	24h	2/5	1/5	1/5	1/5	0/5	5/5	1/5	0/5	0/5	1/5	5/5	0/5	0/5	0/5	5/5		
ATCC 43889 (2.7ctu/g)	24h	5/5	5/5	5/5	4/5	4/5	5/5	5/5	4/5	5/5	4/5	5/5	5/5	5/5	5/5	5/5		
TT7 (1.8 cfu/g)	24h	5/5	5/5	5/5	5/5	2/5	5/5	5/5	5/5	5/5	2/5	5/5	1/5	5/5	5/5	1/5		
Total		19/25	19/25	17/25	16/25	10/25	23/25	20/25	17/25	19/25	15/25	24/25	13/25	17/25	15/25	17/25		
Acid Treated																		
ATCC 35150 (1.8 ctu/g)	24h		5/5	5/5	5/5	5/5		5/5	5/5	5/5	5/5		5/5	5/5	5/5	5/5		
ATCC 43894 (2.0 ctu/g)	24h		5/5	5/5	5/5	5/5		5/5	5/5	5/5	5/5		5/5	5/5	5/5	5/5		
ATCC 43895 (1.8 ctu/g)	24h		5/5	1/5	5/5	1/5		5/5	4/5	5/5	5/5		5/5	5/5	5/5	5/5		
ATCC 43889 (2.7ctu/g)	24h		5/5	5/5	5/5	5/5		5/5	5/5	5/5	5/5		5/5	5/5	5/5	5/5		
TT7 (1.8 cfu/g)	24h		5/5	5/5	5/5	5/5		4/5	4/5	4/5	4/5		5/5	5/5	5/5	5/5		
Total			25/25	21/25	25/25	21/25		24/25	23/25	24/25	24/25		25/25	25/25	25/25	25/25		

and vancomycin (8mg/L) and incubated with shaking at 42°C for 24 h. D etection by real time PCR, and cultural isolation with or without IMS and an acid treatment

Table 5. Detection of E. coli O157:H7 in alfalfa sprouts artificially contaminated at a high level (1-3 cfu/g) with EHEC and enriched in mBPWp with cefsulodin (10mg/L) and vancomvcin (8mg/L) and incubated with shaking at 42°C for 24 h. p etection by real time PCR, and cultural isolation with or without IMS and an acid treatr

E.coli 0157 Strain I (inoculum level)	Incubation	Broth					Pathatrix beads						Dynabeads® M AX E.coli 0157					
	Time	PCR	Cultural isolation	Rainbow	TC-SMAC	R & F	PCR	Cultural isolation	Rainbow	TC-SMAC	R&F	PCR	Cultural isolation	Rainbow	TC-SMAC	R & F		
ATCC 35150 (0.18 cfu/g)	24h	4/5	5/5	4/5	3/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5	3/5	3/5	4/5		
ATCC 43894 (0.2 cfu/g)	24h	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5	4/5	5/5	5/5		
ATCC 43895 (0.18 cfu/g)	24h	5/5	5/5	5/5	4/5	3/5	5/5	4/5	1/5	4/5	3/5	5/5	4/5	1/5	4/5	1/5		
ATCC 43889 (0.27cfu/g)	24h	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5	4/5	5/5	5/5		
TT7 (0.18 cfu/g)	24h	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5	5/5		
Total		24/25	25/25	24/25	22/25	23/25	25/25	24/25	21/25	24/25	23/25	25/25	24/25	17/25	22/25	20/25		
Acid Treated																		
ATCC 35150 (0.18 cfulg)	24h		5/5	5/5	5/5	5/5		5/5	5/5	5/5	5/5		5/5	5/5	5/5	5/5		
ATCC 43894 (0.2 cfu/g)	24h		5/5	5/5	5/5	5/5		5/5	5/5	5/5	4/5		5/5	5/5	5/5	5/5		
ATCC 43895 (0.18 cfu/g)	24h		5/5	5/5	5/5	5/5		5/5	5/5	4/5	2/5		5/5	5/5	5/5	5/5		
ATCC 43889 (0.27cfu/g)	24h		5/5	5/5	5/5	5/5		5/5	5/5	5/5	5/5		5/5	5/5	5/5	5/5		
TT7 (0.18 cfu/g)	24h		5/5	5/5	5/5	5/5		5/5	5/5	5/5	5/5		5/5	5/5	5/5	5/5		
Total			25/25	25/25	25/25	25/25		25/25	25/25	24/25	21/25		25/25	25/25	25/25	25/25		

CONCLUSIONS

• Low levels of EHEC can be reliably detected by real-time PCR in alfalfa sprouts following enrichment in mBPWp with CV added at the onset of incubation, incubating the samples at elevated temperature (42°C) with shaking at 100 rpm throughout the enrichment

Isolation on selective agar can be difficult due to the presence of competing microorganisms on the selective agar plates even in samples where the EHEC are concentrated by immunomagnetic separation.

Cultural recovery of E. coli O157 was easier when the acid treatment was used either directly or with IMS beads.

REFERENCES

 Weagant, S.D. and A.J. Bound. (2001) Evaluation of techniques for enrichment and isolation of *Escherichia coli* 0157147 from artificially contaminated sprouts. Inter. J. Food Microbiol. 71:87-92.
Jinneman, K.C., K.J. Voshthouri, and S.D. Weagant (2003) Multiplex real-time PCR method to identify Shiga Louising and Link Wendakoon, C., S.D. Weagant, C. Carrillo, K.J. Yoshitomi, K.C. Jinneman, R. Zapata, P. Browning and W.M. Fedio. (2008). Optimization of a rapid method for detection of *E*. coli 0157:H71 in alfalla sproats using real time PCR combined with immunomagnetic separation. American Society for Microbiology annual meeting. Boston, MA. 5. Jinneman, K.C., K.J. Yoshitomi, S.D. Weagant, R. Zanata, P. Browning, and w. M. Fedio (2009). Evaluation of a 5. Jinéman, K.J., K.J. Iosintóm, S.J. Weigian, K. Zajara, S. Rowmag, and W.M. Fedio (2009) Evaluation o rapid procedure for detection of E evol (0157:H7) in alfalfa sporous using real-time PCR combined with immunomagenite separation. Institute of Food Technologista snaula meeting. Anaheim, CA. Fedio, W.R. Zapata, K. Yoshibani, K. Jinneman, S. Soteka, and S. Weagant, Detection of very low levels of E. coli O157:H7 in cilantro by immunomagnetic separation, real-time PCR and cultural methods with and without an acid treatment. Institute of Food Technologists annual meeting, Aneheim, Ca (2009).

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